Novel Dysregulated MicroRNAs in Primary Laryngeal Squamous Cell Cancer

Kang Mei Chen, Josena K Stephen, Shaleta Havard, Veena Shah, Glendon Gardner Vanessa G Schweitzer, Maria J Worsham

ABSTRACT

Introduction: MicroRNAs (miRNAs) are endogenous, small, noncoding RNAs of 17 to 25 nucleotides that regulate approximately 30% of human genes. They are differentially expressed in various types of cancers compared with noncancerous tissues, suggesting that they may have crucial roles in tumorigenesis. The objective of this study was to identify laryngeal squamous cell cancer (LSCC)-specific miRNAs.

Materials and methods: A retrospective cohort of 10 LSCC and five normal laryngeal squamous epithelium samples were examined using a global miRNA profiling approach (HTG, Tucson, AZ, USA, 800 human miRNAs plus 10 endogenous control miRNAs). The expression status of selected dysregulated miRNAs that were significantly different from normal were verified by real-time quantitative PCR (qPCR).

Results: Twenty-three of the 800 human miRNAs had significantly different expression levels (p < 0.05) between LSCC and normal tissues. Fifteen of the 23 have not been previously reported in HNSCC and include: miR-663b, miR-663, miR-193b, miR-1291, miR-720, miR-191, miR-1224-3p, miR-214, miR-1285, miR-1207-5p, miR-483-5p, miR-1225-3p, miR-1228, miR-1280 and miR-638. Consistently upregulated miR-31 and miR-193b and differentially expressed miR-663b in LSCC were verified by qPCR.

Conclusion: The 15 novel miRNAs identified in this exploratory study, pending further confirmation and validation, may have clinical utility as LSCC-specific markers.

Keywords: MicroRNAs, Laryngeal squamous cell cancer, Global microRNA profiling, qPCR validation.

How to cite this article: Chen KM, Stephen JK, Havard S Shah V, Gardner G, Schweitzer VG, Worsham MJ. Novel Dysregulated MicroRNAs in Primary Laryngeal Squamous Cell Cancer. Int J Head Neck Surg 2012;3(2):76-81.

Source of support: Nil

Conflict of interest: None

INTRODUCTION

MicroRNAs (miRNAs) are endogenous, small, noncoding RNAs of 17 to 25 nucleotides that are thought to regulate approximately 30% of human genes.¹⁻³ They are involved in regulating target gene expression through imperfect base pairing with the 3'-untranslated region (3'-UTR) of target mRNAs of protein-coding genes, leading to the cleavage of homologous mRNA or translational inhibition. They are differentially expressed in various types of cancers compared with noncancerous tissues, suggesting that they may have crucial roles in tumorigenesis.⁴⁻⁸

Quite a few miRNAs have been linked to head and neck squamous cell carcinoma (HNSCC) based on their differential expression in tumors. miR-21, a commonly dysregulated miRNA in cancer, is frequently upregulated in HNSCC.⁹⁻¹⁴ Progression of oral cancers has been significantly associated with miR-345, -21 and -181b.12 Significantly low levels of miR-125a and miR-200a have been detected in the saliva of oral cancer patients¹⁵ while high levels of miR-184 have been detected in the plasma of tongue SCC.¹⁶ Altered expression of specific miRNAs are beginning to provide much needed insights into tumorigenesis mechanisms of abnormal cell-cycle regulation, evasion of apoptosis, reduced response to antigrowth signals and epithelial-mesenchymal transition (EMT).¹⁷ These reports strongly support the potential utility of miRNAs as diagnostic biomarkers in HNSCC. The objective of this pilot study was to identify laryngeal squamous cell carcinoma (LSCC)-specific miRNAs using a global discovery approach.

MATERIALS AND METHODS

Cohort

The retrospective pilot cohort comprised 10 primary LSCC cases (cancer cohort) and five normal laryngeal squamous epithelium tissues (control cohort). This study was approved by the Henry Ford Health System Institutional Review Board Committee.

RNA Extraction

miRNA from tumor and normal tissue, present in separate formalin-fixed, paraffin-embedded (FFPE) tissue blocks was extracted using the High pure miRNA Isolation Kit (Roche). Final dilution of total RNA used for reverse transcription was 20 ng/ul.

Quantitative Nuclease Protection miRNA Microarray Assay

Global miRNA profiling using quantitative nuclear protection (qNPA) miRNA microarray assays was provided by High Throughput Genomics Inc. (HTG, Tucson, AZ, USA). This assay interrogates a panel of 800 human miRNAs plus 10 endogenous control miRNAs simultaneously.¹⁸ We provided two curls of 5 μ m tissue sections in tubes for each of the 15 cases.

HTG Data Analysis

Each microarray has two elements (replicates) for each transcript (A1.1 and A1.2, for example); therefore the total number of data points for a single transcript is 2. Data is normalized to the total signal for each microarray. A particular transcript is considered expressed, if the signal is more than three standard deviations above the background signal as determined by the *ANT* gene.

Significantly differently expressing miRNAs between normal and tumor samples were obtained using the Student's t-test. Significantly (p < 0.05) upregulated and downregulated miRNAs detected in the LSCC group (as compared to the normal group) by the qNPA assay are listed in Table 1.

Table 1: Significantly upregulated and downregulated miRNAs						
miRNA	Ratio	p-value	Chromosome location			
Upregulated	6 67	0.000	2			

hsa-miR-663b ¹	6.67	0.000	2
hsa-miR-320a	4.81	0.000	8p21.3
hsa-miR-320b	4.32	0.000	1
hsa-miR-663	3.38	0.000	20p11.1
hsa-miR-193b [†]	10.83	0.001	16p13.12
hsa-miR-92a	5.72	0.001	13q31.3/Xq26.2
hsa-miR-1291	3.98	0.001	12
hsa-miR-16	3.97	0.003	13q14.3/3q26.1
hsa-miR-720	2.07	0.004	3
hsa-miR-27a	5.11	0.005	19p13.13
hsa-let-7f	2.89	0.006	9q22.32/Xp11.22
hsa-miR-191	3.30	0.009	3p21.31
hsa-miR-1224-3p	3.19	0.009	3q27.1
hsa-miR-31 [†]	6.16	0.010	9p21.3
hsa-miR-214	3.15	0.010	1q24.3
hsa-miR-1285	2.23	0.010	7/2
Downregulated			
0	0.61	0.000	8
hsa-miR-1207-5p	0.61		-
hsa-miR-483-5p	0.62	0.000	11p15.5
hsa-miR-1280	0.76	0.000	3
hsa-miR-1228	0.76	0.001	12
hsa-miR-1225-3p	0.76	0.002	16p13.3
hsa-miR-296-5p	0.75	0.008	20q13.32
hsa-miR-638	0.83	0.035	19p13.2

[†]Verified by qRT-PCR

Bolded: Previously unreported miRNAs

Five miRNAs with the lowest p-values (three upregulated: miR-663b, miR-193b, and miR-31; two downregulated: miR-923 and miR-1826) were selected for further verification by quantitative real-time PCR.

TaqMan MicroRNA Reverse Transcription

Reverse transcription was performed using TaqMan MicroRNA Reverse Transcription Kit (ABI) which was used in conjunction with the TaqMan MicroRNA Individual Assays (ABI) containing the individual reverse transcription

primers. The manufacturer's prescribed protocol was followed for both kits. Final volume for 1 reaction is 7.5 ul (3.5 ul RT master mix, 2.5 ul total RNA, and 1.5 ul RT primer).

Real-Time Quantitative PCR (qPCR)

Real-time quantitative PCR (qPCR) approach was employed to verify expression status of selected miRNAs that were significantly different from normal controls as proof-ofprinciple. This was performed using TaqMan Universal Master Mix II (no UNG). Manufacturer's protocol was followed for microRNA individual assays. Each sample was run in duplicate.

qPCR Data Analysis

miRNA expression in the normal and tumor groups was measured using $^{\Delta\Delta}$ Ct relative quantization. miRNAs with a value <0 were considered as downregulated and >0 were upregulated (Fig. 1).

RESULTS

All five normal samples and nine of 10 LSCC cases were part of the analyses (one tumor sample indicated outlier values and was excluded). Twenty-three of the 800 human miRNAs were differentially expressed (p < 0.05) between LSCC and normal tissues (Table 1), of which 16 were upregulated and seven were downregulated.

Fifteen of the 23 dysregulated miRNAs have not been previously reported in HNSCC and include: miR-663b,

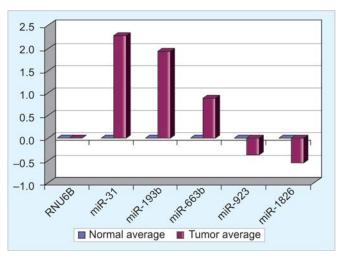


Fig. 1: qPCR data—miRNA expression levels in normal and tumor groups

RNU6B, the internal control miRNA, is not expressed in normal or tumor samples. miR-31, miR-193b and miR-663b are upregulated, whereas miR-923 and miR-1826 were downregulated in the tumor samples. miR-923 and miR-1826 are now listed as fragments in recent updates of the miRNA database and, therefore, not considered as miRNAs miR-663, miR-193b, miR-1291, miR-720, miR-191, miR-1224-3p (3' arm of miR-1224 hairpin), miR-214, miR-1285, miR-1207-5p (5' arm of miR-1207 hairpin), miR-483-5p, miR-1225-3p, miR-1228, miR-1280 and miR-638 (highlighted in bold in Table 1).

Selected miRNAs, miR-923, miR-1826, miR-663b, miR-193b and miR-31, with p-values 0.000, 0.000, 0.000, 0.001 and 0.01 respectively, were verified by qPCR and were concordant with microarray data. miR-31 and miR-193b were consistently upregulated (9/9), and miR-663b was differentially regulated (7/9 upregulated and 2/9 down-regulated) in LSCC (Table 2). miR-923 and miR-1826 (not shown in Table 1) showed downregulation in 6/9 and upregulation in 3/9 samples; however, these are now listed as fragments in recent updates of the miRNA database and no longer regarded as legitimate miRNAs.

DISCUSSION

miRNAs, though lurking behind the scenes, are now being viewed less as molecular noise and more as exotic players with increasing prominence in theories about cancer.¹⁹ By binding to mRNA, miRNA can silence or modulate mRNA function. Expression profiling of miRNAs has shown that some miRNAs are upregulated or down-regulated in cancer, suggesting that it is important to understand the specific roles miRNAs may have in cancer.

There are two types of cancer-related miRNAs: Oncogenic (i.e. miR-155, miR-21)⁸ or tumor suppressor miRNAs (i.e. miR-15a and let-7 family).^{20,21} In HNSCC, miRNA profiling has been performed in oral, oropharyngeal, tongue, laryngeal and thyroid cancers. A study by Hui et al²² found no distinct differences in the global miRNA profiles between squamous cell cancers arising from the larynx, oropharynx or hypopharynx with the exception of miR-133b which was more highly expressed in laryngeal *vs* the other two subsites. They found about a third of the miRNAs examined in their HNSCC samples to be dysregulated. Because several of the differentially expressed miRNAs in their study were in frequently amplified or deleted regions, genomic amplifications or deletions were offered as one possible mechanism for abnormal miRNA expression in HNSCC.

We found 23 significantly differentially expressed miRNAs for many of which, information is limited to chromosomal locations (Table 1). Of the 23 significantly differentially expressed miRNAs, eight have been reported previously in HNSCC. These include miR-320a, miR-320b, miR-92a, miR-16, miR-27a, let-7f, miR-31 and miR-296-5p.

miR-27a is significantly upregulated in HNSCC cell lines and its expression is correspondingly reduced by using knock down anti sense approaches.²² It is considered a noncausal HNSCC miRNA as its manipulation in vitro does not have an effect on phenotype.¹⁷ miR-92a, previously named miR-92 in the miRNA database, has two precursor sequences: miR-92a-1 and miR-92a-2.23 miR-92a-1 is located on the oncogenic miR-17-92 polycistron of chromosome 13q31.3, a locus that harbors quite a few upregulated miRNAs.²² mir-92a-2, located on chromosome Xq26.2, belongs to the miR-106a-92 cluster and is also overexpressed in HNSCC.²² miR-16 is commonly downregulated due to deletion of the 13q14 region in chronic lymphocytic leukemia;^{24,25} however, it is significantly upregulated in HNSCC.²² The let-7 family of miRNA is known as a tumor suppressor miRNA that functions through inhibiting oncogenic mRNAs, such as RASA1, MYC, HMGA2.^{21,26,27} The let-7 family members are commonly downregulated in HNSCC except for let-7i which is upregulated.^{10,22,28} Low levels of both let-7d and miR-205 have been reported to be significantly related to poor survival in primary HNSCC.¹¹

miR-31, dysregulated in many cancers,²⁹⁻³² appears to favor upregulation in HNSCC¹⁴ and oral squamous cell carcinoma (OSCC)^{6,16} cell lines. Its function in tumorigenesis is unclear; however, Liu et al³³ found miR-31 to be

Table 2: qPCR confirmation of qNPA assay								
	miR-31	miR-193b	miR-663b	miR-923	miR-1826			
LT-ID6	Up	Up	Up	Down	Down			
LT-ID8	Up	Up	Down	Down	Up			
LT-ID9	Up	Up	Up	Up	Up			
LT-ID10	Up	Up	Up	Down	Down			
LT-ID11	Up	Up	Up	Down	Down			
LT-ID12	Up	Up	Up	Down	Down			
LT-ID13	Up	Up	Up	Up	Up			
LT-ID14	Up	Up	Up	Up	Down			
LT-ID15	Up	Up	Down	Down	Down			
	9/9 up	9/9 up	7/9 up	6/9 down	6/9 down			
			2/9 down	3/9 up	3/9 up			

 $^{\Delta\Delta}C_t$ relative quantization (tumor $^{\Delta}Ct$ —normal $^{\Delta}Ct$) used to determine up or downregulation

significantly elevated in the plasma of OSCC patients, which was remarkably reduced following surgery. This suggests that miR-31 may be an oncogenic miRNA. Its detection in plasma could be clinically useful as a noninvasive diagnostic approach. miR-296 is downregulated in OSCC.⁶

Several studies have reported significant differential expression of miR-100, miR-125b and miR-375 in HNSCC,^{5,13,14,16,34} none of which reached significance in this study.

Of the 23 significantly differentially expressed miRNAs, 15 have not been previously reported in HNSCC and include: miR-663b, miR-663, miR-193b, miR-1291, miR-720, miR-191, miR-1224-3p, miR-214, miR-1285, miR-1207-5p, miR-483-5p, miR-1225-3p, miR-1228, miR-1280 and miR-638.

Dysregulation of miR-193b is thought to influence melanoma development³⁵ via its role of cell proliferation repression and regulation of CCND1 expression.³⁵ In prostate cancer, miR-193b can present as an epigenetically silenced putative tumor suppressor.³⁶ It has also been detected in cervical cancer cell lines.³⁷ The role of miR-663b reported in the development of human leukemias²⁴ and of miR-663 in human colorectal cells³⁸ is not known.

miR-191 is upregulated in hepatocellular carcinoma (HCC) and affects the *TGF*- β and *MAPK* pathways, which play an important role in HCC tumorigenesis.^{39,40} Inhibition of miR-191 by 2-*O*-metoxyethyl (MOE) anti-miR was found to decrease cell proliferation and induce apoptosis *in vitro* with significant reduction of tumor mass in an *in vivo* mouse model of HCC. Therefore, in HCC, miR-191 may signal a potential therapeutic target.

miR-1224-3p has been identified as a mammalian mirtron which is a short hairpin intron that is a precursor for miRNA biogenesis.⁴¹ Mirtrons were originally found in invertebrates but are now known to exist in mammals also. miR-214 has been detected in melanomas, cervical and ovarian cancers.⁴²⁻⁴⁴ In melanomas, miR-214 suppresses TFAP2C leading to tumor progression.⁴² miR-1285 has been shown to inhibit expression of TP53.45 miR-1207-5p is expressed abundantly in colon cancer cell lines along with miR-1207-3p, and Northern blot data suggested a common promoter and transcriptional regulatory unit making these likely complimentary overlapping miRNAs.⁴⁶ miR-483-5p is significantly upregulated in malignant adrenocortical carcinoma (ACC) as compared with benign tumors.⁴⁷ Its expression can accurately distinguish tumors as benign or malignant.

miR-638 has been detected in human colorectal cells³⁸ and in gastric cancer,⁴⁸ where it is significantly downregulated. It is present in a stable form in human plasma. Tanaka et al.⁴⁹ found that the ratio of miR-92a/

miR-638 in plasma is useful for distinguishing leukemia patients from healthy individuals.

CONCLUSION

The human miRNA database continues to get fine tuned with respect to new additions as well as exclusions. This pilot study identified previously unreported miRNAs in LSCC that upon further investigation in larger studies may have clinical utility in LSCC. Our 23 aberrantly expressed miRNAs in LSCC, including the 15 unreported in HNSCC, require further examination with subsequent validation in larger cohorts for clinical relevance as LSCC-specific markers.

ACKNOWLEDGMENT

Drs Chen and Worsham had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. This study was supported by R01 NIH DE 15990 (Dr Worsham).

REFERENCES

- 1. Bartel DP. MicroRNAs: Genomics, biogenesis, mechanism and function. Cell 2004;116:281-97.
- 2. Bartel DP. MicroRNAs: Target recognition and regulatory functions. Cell 2009;136:215-33.
- 3. Lee YS, Dutta A. MicroRNAs in cancer. Annu Rev Pathol 2009; 4:199-227.
- Christensen BC, Moyer BJ, Avissar M, et al. A let-7 microRNAbinding site polymorphism in the KRAS 3' UTR is associated with reduced survival in oral cancers. Carcinogenesis 2009; 30:1003-07.
- Henson BJ, Bhattacharjee S, O'Dee DM, Feingold E, Gollin SM. Decreased expression of miR-125b and miR-100 in oral cancer cells contributes to malignancy. Genes Chromosomes Cancer 2009;48:569-82.
- Kozaki K, Imoto I, Mogi S, Omura K, Inazawa J. Exploration of tumor-suppressive microRNAs silenced by DNA hypermethylation in oral cancer. Cancer Res 2008;68:2094-105.
- 7. Calin GA. MicroRNAs and cancer: What we know and what we still have to learn. Genome Med 2009;1:78.
- Garzon R, Calin GA, Croce CM. MicroRNAs in Cancer. Annu Rev Med 2009;60:167-79.
- Li J, Huang H, Sun L, et al. MiR-21 indicates poor prognosis in tongue squamous cell carcinomas as an apoptosis inhibitor. Clin Cancer Res 2009;15:3998-4008.
- Chang SS, Jiang WW, Smith I, et al. MicroRNA alterations in head and neck squamous cell carcinoma. Int J Cancer 2008; 123:2791-97.
- Childs G, Fazzari M, Kung G, et al. Low-level expression of microRNAs let-7d and miR-205 are prognostic markers of head and neck squamous cell carcinoma. Am J Pathol 2009;174: 736-45.
- Cervigne NK, Reis PP, Machado J, et al. Identification of a microRNA signature associated with progression of leukoplakia to oral carcinoma. Hum Mol Genet 2009;18:4818-29.

- Avissar M, Christensen BC, Kelsey KT, Marsit CJ. MicroRNA expression ratio is predictive of head and neck squamous cell carcinoma. Clin Cancer Res 2009;15:2850-55.
- Tran N, McLean T, Zhang X, et al. MicroRNA expression profiles in head and neck cancer cell lines. Biochem Biophys Res Commun 2007;358:12-17.
- Park NJ, Zhou H, Elashoff D, et al. Salivary microRNA: Discovery, characterization and clinical utility for oral cancer detection. Clin Cancer Res 2009;15:5473-77.
- Wong TS, Liu XB, Wong BY, Ng RW, Yuen AP, Wei WI. Mature miR-184 as potential oncogenic microRNA of squamous cell carcinoma of tongue. Clin Cancer Res 2008; 14:2588-92.
- 17. Babu JM, Prathibha R, Jijith VS, Hariharan R, Pillai MR. A miR-centric view of head and neck cancers. Biochim Biophys Acta 2011;1816:67-72.
- Martel RR, Botros IW, Rounseville MP, et al. Multiplexed screening assay for mRNA combining nuclease protection with luminescent array detection. Assay Drug Dev Technol 2002; 1:61-71.
- 19. Johnson G. Cancer's Secrets Come Into Sharper Focus New York Times, 2011.
- 20. Cimmino A, Calin GA, Fabbri M, et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. Proc Natl Acad Sci USA 2005;102:13944-49.
- 21. Lee YS, Dutta A. The tumor suppressor microRNA let-7 represses the HMGA2 oncogene. Genes Dev 2007; 21:1025-30.
- 22. Hui AB, Lenarduzzi M, Krushel T, et al. Comprehensive MicroRNA profiling for head and neck squamous cell carcinomas. Clin Cancer Res 2010; 16:1129-39.
- 23. Mourelatos Z, Dostie J, Paushkin S, et al. miRNPs: A novel class of ribonucleoproteins containing numerous microRNAs. Genes Dev 2002;16:720-28.
- 24. Takada S, Yamashita Y, Berezikov E, et al. MicroRNA expression profiles of human leukemias. Leukemia 2008; 22:1274-78.
- Calin GA, Dumitru CD, Shimizu M, et al. Frequent deletions and down-regulation of microRNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. Proc Natl Acad Sci USA 2002;99:15524-29.
- 26. Johnson SM, Grosshans H, Shingara J, et al. RAS is regulated by the let-7 microRNA family. Cell 2005;120:635-47.
- 27. Sampson VB, Rong NH, Han J, et al. MicroRNA let-7a downregulates MYC and reverts MYC-induced growth in Burkitt lymphoma cells. Cancer Res 2007;67:9762-70.
- 28. Ramdas L, Giri U, Ashorn CL, et al. miRNA expression profiles in head and neck squamous cell carcinoma and adjacent normal tissue. Head Neck 2009;31:642-54.
- 29. Wong QW, Lung RW, Law PT, et al. MicroRNA-223 is commonly repressed in hepatocellular carcinoma and potentiates expression of Stathmin. Gastroenterology 2008;135:257-69.
- Slaby O, Svoboda M, Fabian P, et al. Altered expression of miR-21, miR-31, miR-143 and miR-145 is related to clinicopathologic features of colorectal cancer. Oncology 2007; 72:397-402.
- Zhang Y, Guo J, Li D, et al. Down-regulation of miR-31 expression in gastric cancer tissues and its clinical significance. Med Oncol 2010;27:685-89.
- 32. Schaefer A, Jung M, Mollenkopf HJ, et al. Diagnostic and prognostic implications of microRNA profiling in prostate carcinoma. Int J Cancer 2010;126:1166-76.

- 33. Liu CJ, Kao SY, Tu HF, Tsai MM, Chang KW, Lin SC. Increase of microRNA miR-31 level in plasma could be a potential marker of oral cancer. Oral Dis 2010;16:360-64.
- Yu T, Wang XY, Gong RG, et al. The expression profile of microRNAs in a model of 7,12-dimethyl-benz[a]anthranceinduced oral carcinogenesis in Syrian hamster. J Exp Clin Cancer Res 2009;28:64.
- Chen J, Feilotter HE, Pare GC, et al. MicroRNA-193b represses cell proliferation and regulates cyclin D1 in melanoma. Am J Pathol 2010;176:2520-29.
- 36. Rauhala HE, Jalava SE, Isotalo J, et al. miR-193b is an epigenetically regulated putative tumor suppressor in prostate cancer. Int J Cancer 2010;127:1363-72.
- Lui WO, Pourmand N, Patterson BK, Fire A. Patterns of known and novel small RNAs in human cervical cancer. Cancer Res 2007;67:6031-43.
- 38. Cummins JM, He Y, Leary RJ, et al. The colorectal microRNAome. Proc Natl Acad Sci USA 2006; 103:3687-92.
- Elyakim E, Sitbon E, Faerman A, et al. hsa-miR-191 is a candidate oncogene target for hepatocellular carcinoma therapy. Cancer Res 2010;70:8077-87.
- Calvisi DF, Pascale RM, Feo F. Dissection of signal transduction pathways as a tool for the development of targeted therapies of hepatocellular carcinoma. Rev Recent Clin Trials 2007;2: 217-36.
- 41. Berezikov E, Chung WJ, Willis J, Cuppen E, Lai EC. Mammalian mirtron genes. Mol Cell 2007;28:328-36.
- Penna E, Orso F, Cimino D, et al. MicroRNA-214 contributes to melanoma tumour progression through suppression of TFAP2C. EMBO J 2011;30:1990-2007.
- 43. Qiang R, Wang F, Shi LY, et al. Plexin-B1 is a target of miR-214 in cervical cancer and promotes the growth and invasion of HeLa cells. Int J Biochem Cell Biol 2011;43:632-41.
- 44. Yin G, Chen R, Alvero AB, et al. TWISTing stemness, inflammation and proliferation of epithelial ovarian cancer cells through MIR199A2/214. Oncogene 2010;29:3545-53.
- 45. Tian S, Huang S, Wu S, Guo W, Li J, He X. MicroRNA-1285 inhibits the expression of p53 by directly targeting its 3' untranslated region. Biochem Biophys Res Commun 2010; 396:435-39.
- Huppi K, Volfovsky N, Runfola T, et al. The identification of microRNAs in a genomically unstable region of human chromosome 8q24. Mol Cancer Res 2008;6:212-21.
- 47. Patterson EE, Holloway AK, Weng J, Fojo T, Kebebew E. MicroRNA profiling of adrenocortical tumors reveals miR-483 as a marker of malignancy. Cancer 2011;117:1630-39.
- 48. Yao Y, Suo AL, Li ZF, et al. MicroRNA profiling of human gastric cancer. Mol Med Report 2009;2:963-70.
- 49. Tanaka M, Oikawa K, Takanashi M, et al. Down-regulation of miR-92 in human plasma is a novel marker for acute leukemia patients. PLoS One 2009; 4:e5532.

ABOUT THE AUTHORS

Kang Mei Chen

Department of Otolaryngology/Head and Neck Research, Henry Ford Hospital, MI, USA

Josena K Stephen

Research Scientist, Department of Otolaryngology/Head and Neck Research, Henry Ford Hospital, MI, USA

Shaleta Havard

Senior Research Assistant, Department of Otolaryngology/Head and Neck Research, Henry Ford Hospital, MI, USA

Veena Shah

Pathologist, Department of Pathology, Henry Ford Hospital, MI USA

Glendon Gardner

Otolaryngologist, Department of Otolaryngology/Head and Neck Surgery, Henry Ford Hospital, MI, USA

Vanessa G Schweitzer

Otolaryngologist, Department of Otolaryngology/Head and Neck Surgery, Henry Ford Hospital, MI, USA

Maria J Worsham (Corresponding Author)

FACMG, Director of Research, Department of Otolaryngology/Head and Neck Research, Henry Ford Hospital, I Ford Place, ID, Detroit MI 48202, USA, Phone: 313-874-3350, Fax: 313-874-1079 e-mail: mworsha1@hfhs.org